

## Supplementary Figures

### Sup Figure 1. H3N1 infection causes AHR and inflammation in WT and NKT cell deficient mice.

**a-c.** 8 wk old BALB/c (**a-b**, n=15 per group) or C57BL/6 (**c**, n=5 per group) mice, treated with influenza A virus (Mem71, H3N1) or control allantoic fluid (mock-infection), were assessed 5, 10 and 15 days post-infection for AHR. (**a,c**) Changes in lung resistance (Penh, enhanced pulse) were measured. \*\*\*p<0.001 compared to mock-infected group. (**b**) Cells in BAL fluid were collected and the numbers of macrophage (Mac), neutrophil (Neu), eosinophil (Eos) and lymphocyte (Lym) were analyzed 10 days after the virus challenge (right panel). \*\*\*p<0.001 compared to mock infected group.

**d.** Representative lung sections stained with H&E from mock or H3N1-infected BALB/c or C57BL/6 mice on day 10.

**e-f.** 8 wk old Wt or *Cd1d*<sup>-/-</sup> mice (n=4 per group) treated with H3N1 or control allantoic fluid, were assessed 5 days post-infection for AHR. Changes in lung resistance (**e**) and cells in BAL fluid were analyzed (**f**). Data are representative of three independent experiments.

### Sup Figure 2. TLR7 agonist induce production of IL-33 in alveolar macrophages.

AM were infected *in vitro* with the indicated amount of poly I:C or R848 or influenza A (H3N1; M.O.I=5; 2X10<sup>5</sup> cells/well, in 96 well plates) for 96 hrs. Supernatants were analyzed for IL-33 by ELISA (eBiosciences). Data are representative of two independent experiments.

### Sup Figure 3. Natural helper cells secrete IL-5.

**a.** Lung cells were taken from H3N1 or mock infected BALB/c or *Rag2*<sup>-/-</sup> mice on day 5 and stimulated with or without PMA + ionomycin for 5 hr. The percentage of lung CD45<sup>+</sup>lin<sup>-</sup>ST2<sup>+</sup>Sca-1<sup>+</sup> cells was assessed by FACS. The second row panels show dot plots for lin<sup>-</sup>ST2<sup>+</sup> cells in lung leukocytes (CD45<sup>+</sup>). After gating on the Lin<sup>-</sup>ST2<sup>+</sup> cells, the cells were analyzed for intracellular IL-5 and Sca-1 expression (third row panels).

**b.** Total RNA from H3N1-infected BALB/c mice lungs was isolated on day 1, 3, 5, 7, or day 14 and analyzed by qRT-PCR for IL-5 mRNA expression.

**c.** The lungs of H3N1 or mock-infected BALB/c mice (n=3 per group) were taken on days 0, 1, 4, and 7 post-infection. The individual lungs were homogenized in 1ml PBS and then assessed for IL-5 protein by ELISA (eBiosciences). \*\*\*p<0.001, compared to mock-infected group. Data are representative of three independent experiments.

**Sup Figure 4. Natural helper cells constitute the major subset of IL-13 secreting cells in the lung after H3N1 infection.**

Lung cells were isolated from mock or H3N1-infected mice, and intracellular IL-13 expression was assessed following stimulation with PMA + ionomycin for 5 hr. The percentage of lung IL-13<sup>+</sup> cells and lin<sup>-</sup>ST2<sup>+</sup> Sca-1<sup>+</sup> cells was assessed by FACS. The second row panels show dot plots for IL-13<sup>+</sup> cells in the live lung cells (first row panels). After gating on the IL-13<sup>+</sup> cells, the cells were analyzed for Lin<sup>-</sup>ST2<sup>+</sup> (blue arrow) and Lin<sup>-</sup>ST2<sup>-</sup> (red arrow, lower panels) cells, and then focused on c-Kit and Sca-1 staining (bottom row panels). Data are representative of three independent experiments.

**Sup Figure 5. IL-13 secretion from natural helper cells and alveolar macrophages.**

**a.** Lung cells were taken from H3N1 or mock infected wt mice on day 1 and further stimulated with PMA + ionomycin for 5 hr. The percentage of lung CD45<sup>+</sup>lin<sup>-</sup>ST2<sup>+</sup>c-Kit<sup>+</sup>Sca-1<sup>+</sup> cells was assessed by FACS. Upper panels show dot plots for Lin<sup>-</sup>ST2<sup>+</sup> and Lin<sup>-</sup>ST2<sup>-</sup> cells in lung leukocytes (CD45<sup>+</sup>). After gating on the Lin<sup>-</sup>ST2<sup>+</sup>c-Kit<sup>+</sup> cells, the cells were analyzed for intracellular IL-13 and Sca-1 expression (lower panels).

**b.** Lung cells from **(a)** were assessed by FACS for the percentage of lung interstitial macrophage (IM; F4/80<sup>+</sup>CD11c<sup>-</sup>), alveolar macrophage (AM; F4/80<sup>+</sup>CD11c<sup>+</sup>) and dendritic cells (DC; F4/80<sup>-</sup>CD11c<sup>+</sup>) in lung leukocytes (CD45<sup>+</sup>). Intracellular IL-13 expression and the absolute cell number were further analyzed by gating on these subsets.

**c.** Alveolar macrophages (AM), bone marrow-derived dendritic cells (BMDC) or a mouse lung epithelial cell line (MLE) were infected with H3N1 (M.O.I=5; upper panel); alveolar macrophages (AM) were infected with H3N1 (M.O.I=5; 0.5; 0.05; lower panel) for 24 hr *in vitro*. Total RNA was extracted from the cells and analyzed by qRT-PCR for IL-13 mRNA expression. \*p<0.05, \*\*\*p<0.001, compared to mock-infected group.

**d.** Lung cells were taken from H3N1 or mock infected BALB/c mice on day 5 and further stimulated with PMA + ionomycin for 5 hr. The percentage of lung T cells (TCRβ<sup>+</sup>) among lung leukocytes (CD45<sup>+</sup>) (upper panels), and intracellular IL-13 and IFN-γ expression in CD4<sup>+</sup> and CD8<sup>+</sup> T cells (middle and lower panels) were assessed by FACS. Data are representative of three independent experiments.

**Sup Figure 6. *Ifnγ*<sup>-/-</sup> mice develop robust H3N1-induced AHR.**

**a.** Total RNA from H3N1-infected BALB/c mice lungs was isolated on day 1, 3, 6, 9 or day 15 and analyzed by qRT-PCR for IFN-γ mRNA expression.

**b-e.** 8 wk old Wt, *Tbx21*<sup>-/-</sup> or *Ifn* $\gamma$ <sup>-/-</sup> mice (a-b, n=5-6 per group), treated with H3N1 or mock infected with allantoic fluid, were assessed 5 days post-infection for AHR. **(b, d)** Changes in lung resistance ( $R_L$ ) as a function of methacholine dose are reported. **(c, e)** Cells in BAL fluid were collected and analyzed 5 days after virus challenge. Data are representative of three independent experiments.

**Sup Figure 7. Natural helper cells do not produce IL-17 after H3N1 infection.**

Lung cells were isolated from mock or H3N1-infected mice, and intracellular IL-17 expression was assessed following by stimulation with PMA + ionomycin for 5 hr. The percentage of CD45<sup>+</sup>Lin<sup>-</sup>ST2<sup>+</sup> cells was assessed by FACS. After gating on the CD45<sup>+</sup>Lin<sup>-</sup>ST2<sup>+</sup> cells (top panels), the cells were analyzed for intracellular IL-17 and Sca-1 expression (bottom panels). Data are representative of three independent experiments.

**Sup Figure 8. Kinetics of viral clearance in mice.**

The lungs of H3N1-infected Wt, *Rag2*<sup>-/-</sup>, *Il13*<sup>-/-</sup> and *Il1rl1(St2)*<sup>-/-</sup> mice were taken on the indicated days after infection and assessed for influenza virus by qRT-PCR. The data are presented as relative PFU/lung on a log scale. Data are representative of two independent experiments.

**Sup Figure 9. Anti-Thy1.2 (CD90) mAb treatment of *Rag2*<sup>-/-</sup> mice depletes Natural Helper cells.**

**a.** Lung cells were taken from H3N1 or mock-infected wild-type mice on day 1 and the percentage of lung CD45<sup>+</sup>Lin<sup>-</sup>ST2<sup>+</sup> Sca-1<sup>+</sup> cells was assessed by FACS.

The upper panels show dot plots for Lin<sup>+</sup>ST2<sup>+</sup> and Lin<sup>-</sup>ST2<sup>+</sup> cells in lung leukocytes (CD45<sup>+</sup>). After gating on the Lin<sup>-</sup>ST2<sup>+</sup>c-Kit<sup>+</sup> cells, the cells were analyzed for Thy1.2 and Sca-1 expression (lower panels). The results indicate that H3N1 infection increases the number of Sca-1<sup>+</sup> natural helper cells in the lung. Data are representative of three independent experiments.

**b.** 8 wk-old *Rag2*<sup>-/-</sup> mice were treated with three injections of anti-Thy1.2 mAb (clone 30-H12; 0.5 mg, day -3,0 day 3), and were then infected with H3N1 virus on day 0. The lung cells were taken from H3N1 infected wt mice on day 5 and the percentage of lung CD45<sup>+</sup>Lin<sup>-</sup>ST2<sup>+</sup>Sca-1<sup>+</sup> cells was assessed by FACS. The lower panels show dot plots for Lin<sup>+</sup>ST2<sup>+</sup> and Lin<sup>-</sup>ST2<sup>+</sup> cells in lung leukocytes (CD45<sup>+</sup>) after Thy1.2 treatment, and indicate that anti-Th1.2 mAb treatment depleted the Thy1.2<sup>+</sup>Lin<sup>-</sup>ST2<sup>+</sup> natural helper cells.

**c.** protocol for adoptive transfer of natural helper cells to *Il13*<sup>-/-</sup> recipients.

**Sup Figure 10. *In vitro* stimulation of natural helper cells induced IL-5 and IL-13 secretions.**

**a.** Lung natural helper cells ( $\text{lin}^-\text{ST2}^+$ ) were isolated from the donors that received IL-33 ( $1\mu\text{g}$ , i.n.), and the purity of  $\text{lin}^-\text{ST2}^+$  cells were assessed by FACS after sorting.

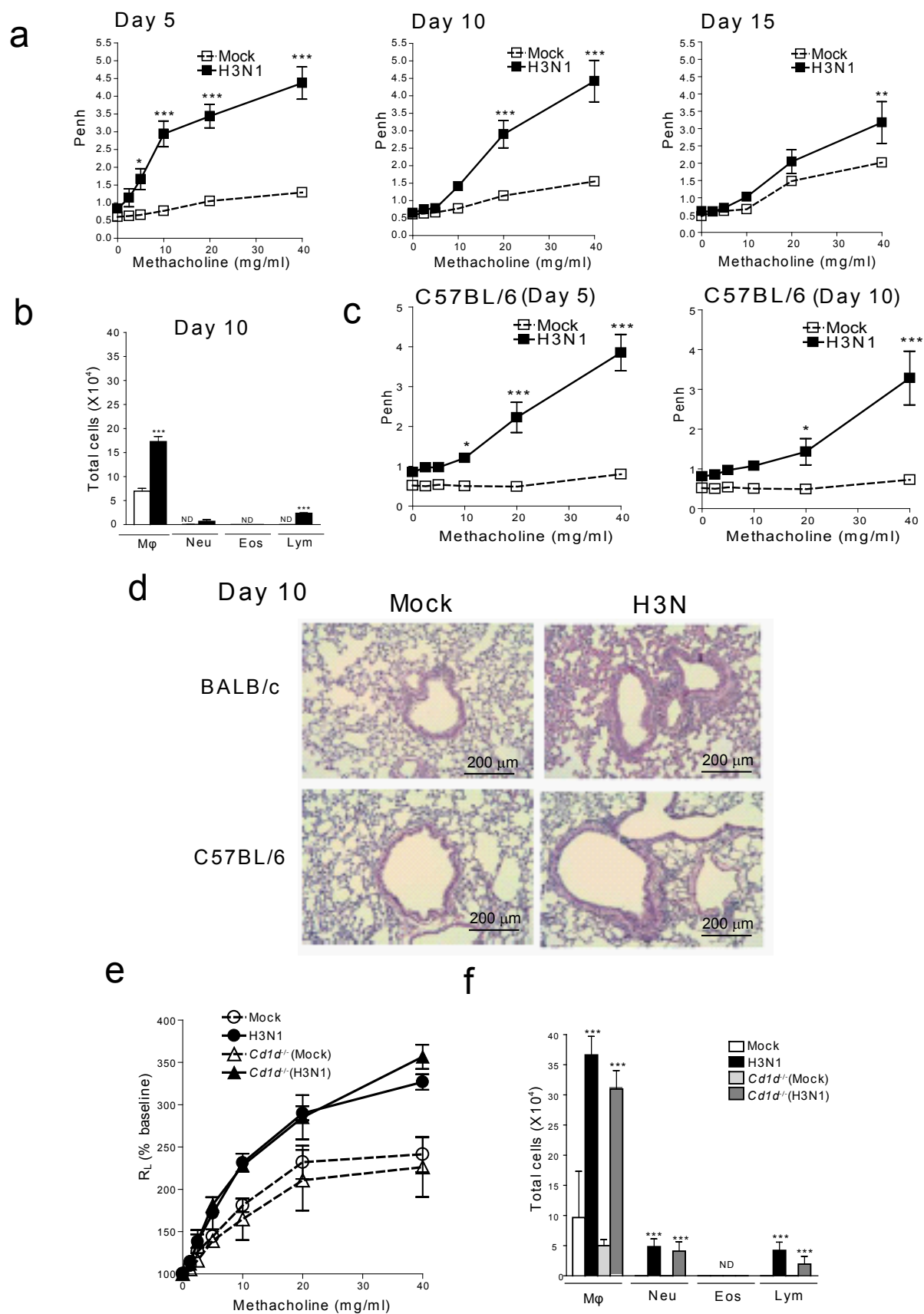
**b-c.** Lung natural helper cells ( $\text{lin}^-\text{ST2}^+$ ) were isolated from the naïve *Rag2*<sup>-/-</sup> donors ( $4 \times 10^4$  cells/well, 96 well plates) were cultured with 50 ng/ml IL-2, 100 ng/ml IL-33 or IL-2 plus IL-33 for 24hrs to 6 days *in vitro*. PMA + ionomycin stimulation was used as a positive control and no cytokine (IL-2 (-)) was used as negative control. Total RNA was extracted from the cells after 24hrs culture, and analyzed by qRT-PCR for IL-5 and IL-13 mRNA expression. Supernatants from triplicate wells were collected on day 1, day 4 and day 6, and then assessed for IL-5 and IL-13 protein by ELISA.

**d.** After 4 days of culture, the cells were harvested and analyzed by FACS for intracellular cytokine expression following stimulation with PMA + ionomycin for 5 hr. The percentage of  $\text{CD45}^+\text{lin}^-\text{ST2}^+$  cells in the live cell gate was assessed by FACS. After gating on the  $\text{CD45}^+\text{lin}^-\text{ST2}^+$  cells (top panels), the cells were analyzed for intracellular IL-5, IL-13, IFN- $\gamma$ , IL-4 and IL-17, compared with respective isotype control. Data are representative of three independent experiments.

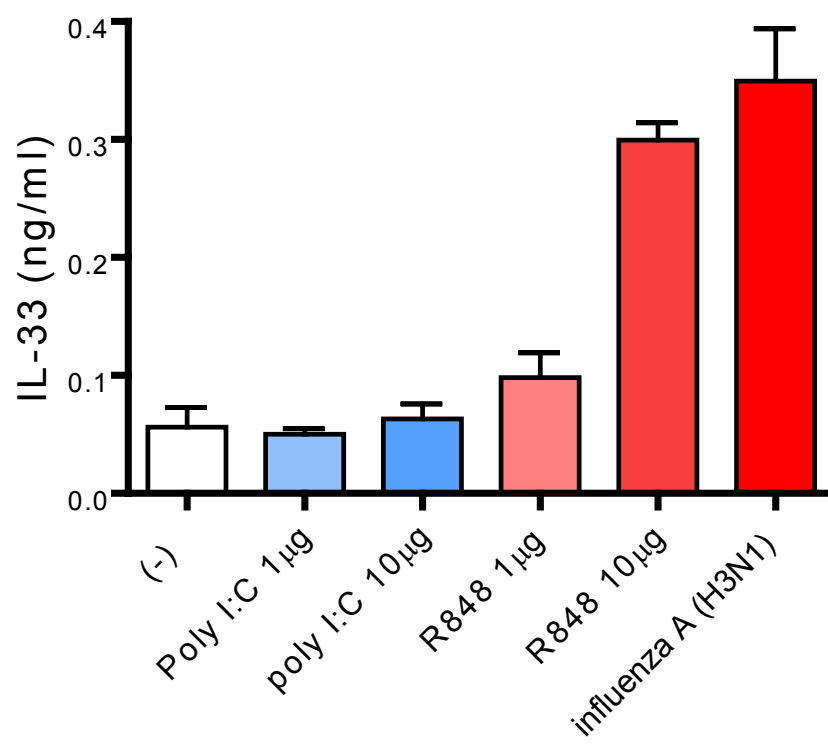
**Sup Figure 11. Schematic showing a macrophage-natural helper cell immune axis leading to acute AHR after influenza virus infection.**

Influenza A virus infects and activates alveolar macrophages (AM), interstitial macrophages (IM), dendritic cells (DC), or lung epithelial cells, resulting in the secretion of IL-33 within 24hr of virus infection. IL-33 production in turn activates natural helper cells (nuocytes) through ST2 receptors. This interaction leads to IL-13 production in natural helper cells (nuocytes) 5-6 days after infection. The IL-13 then drives mucus secretion by airway epithelial cells and the development of AHR.

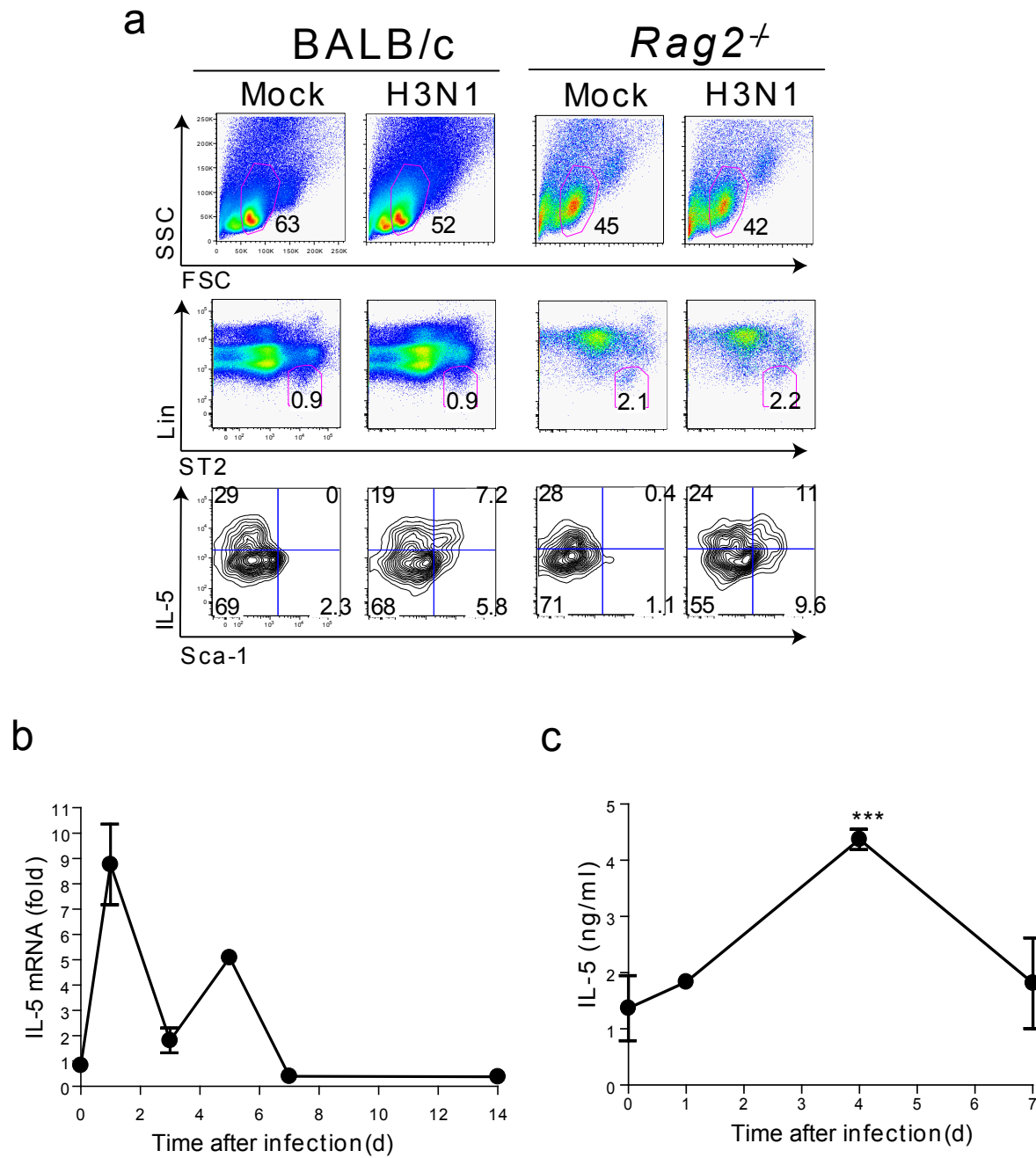
# Sup Fig 1



Sup Fig 2

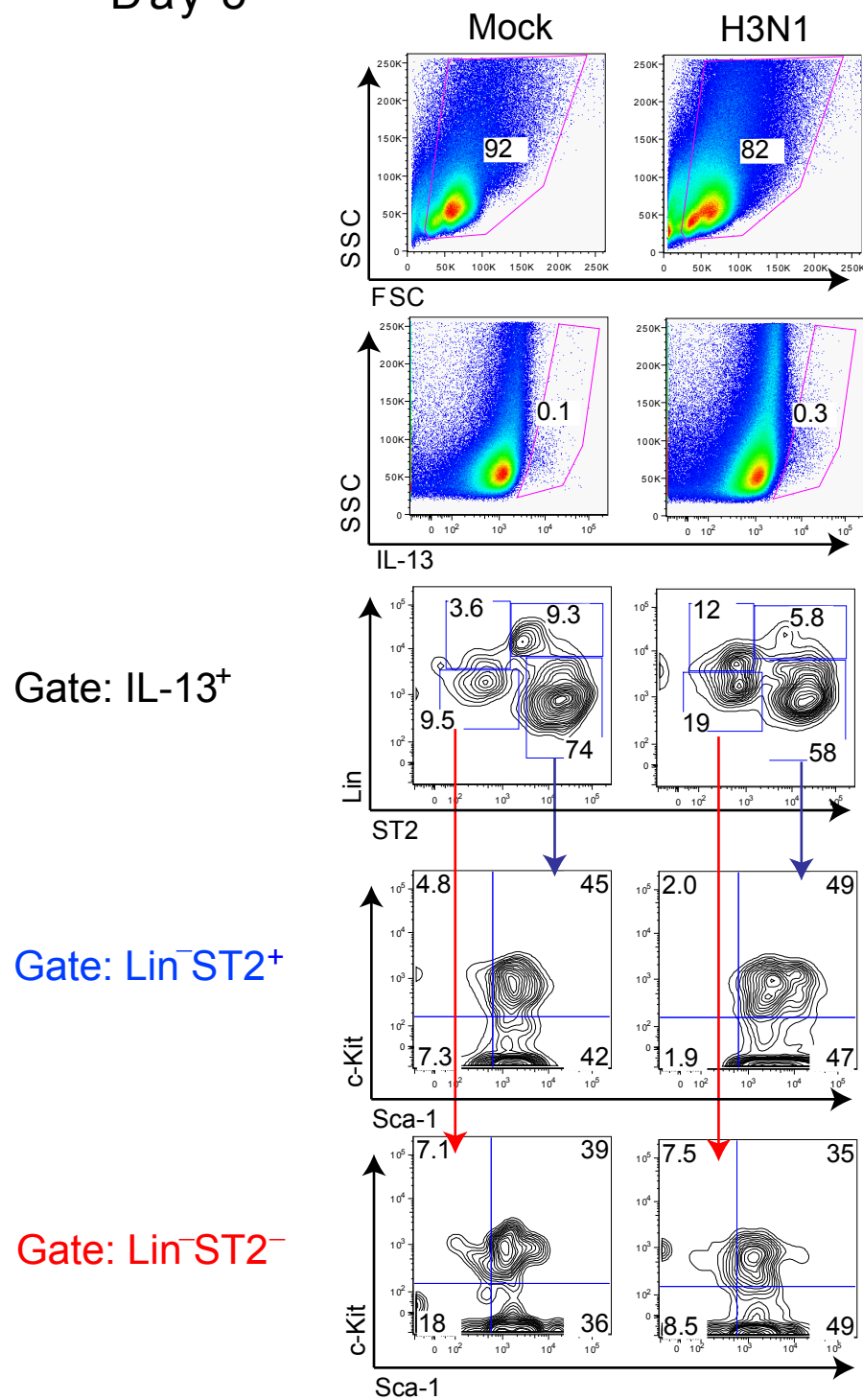


# Sup Fig 3



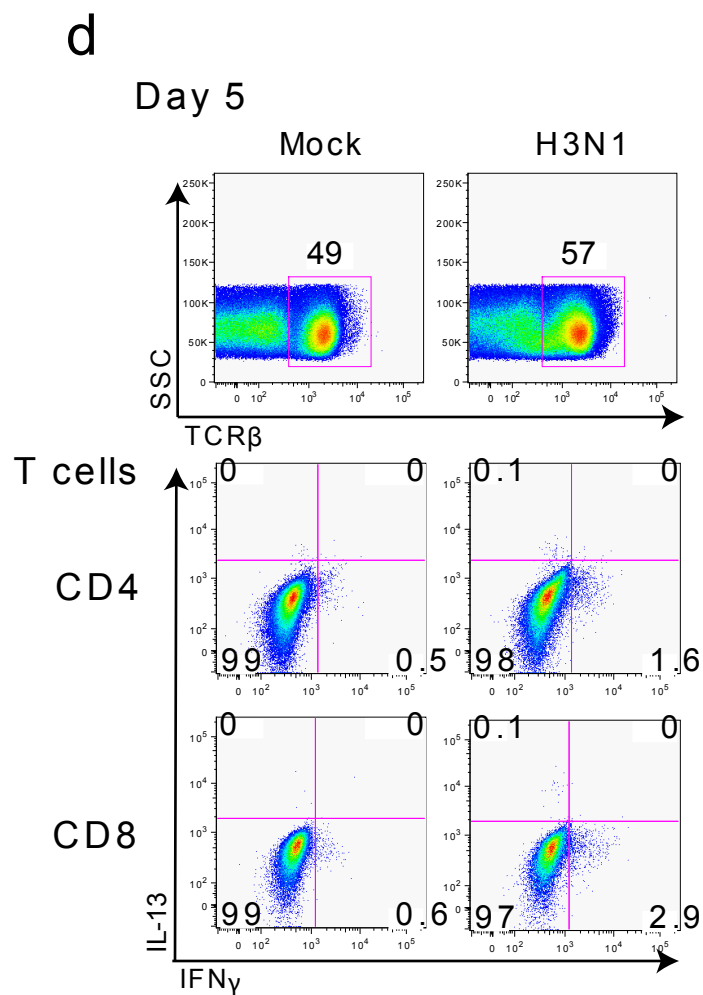
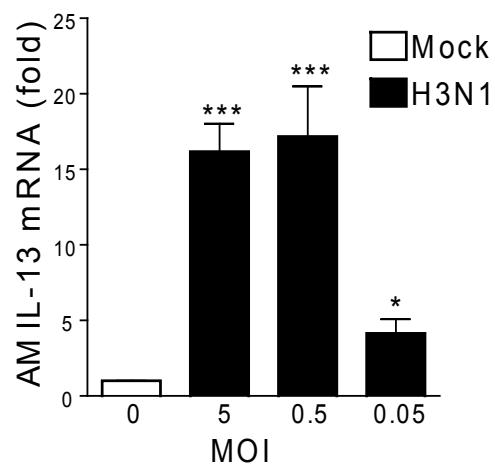
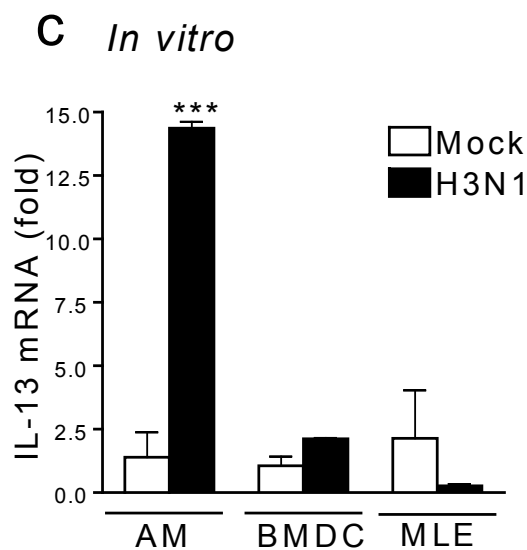
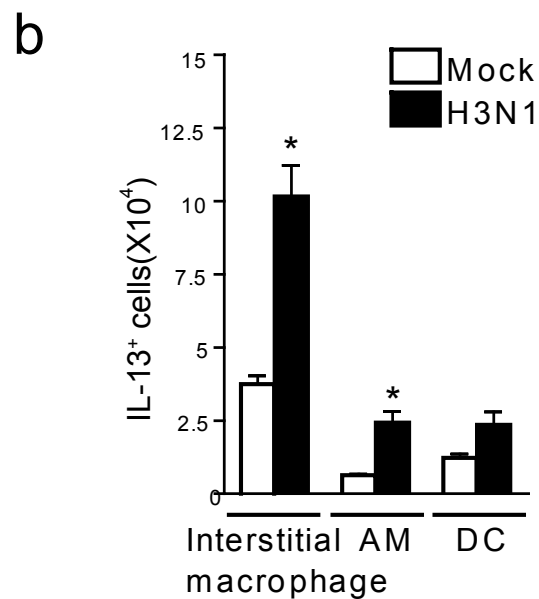
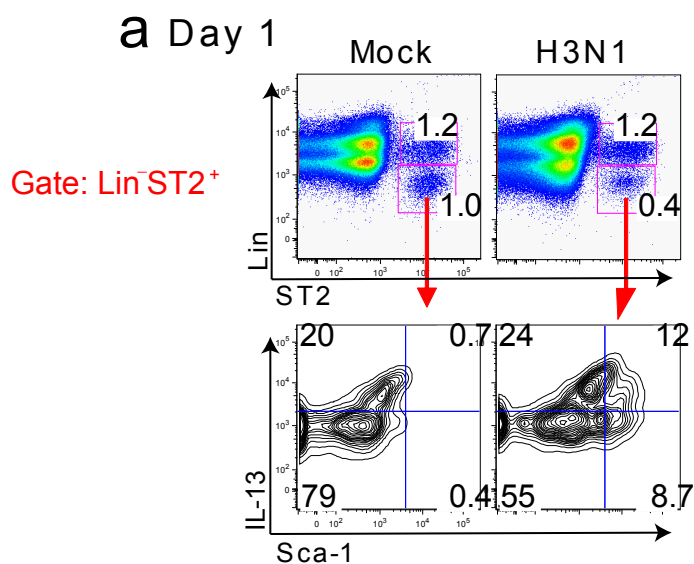
Sup Fig 4

Day 5

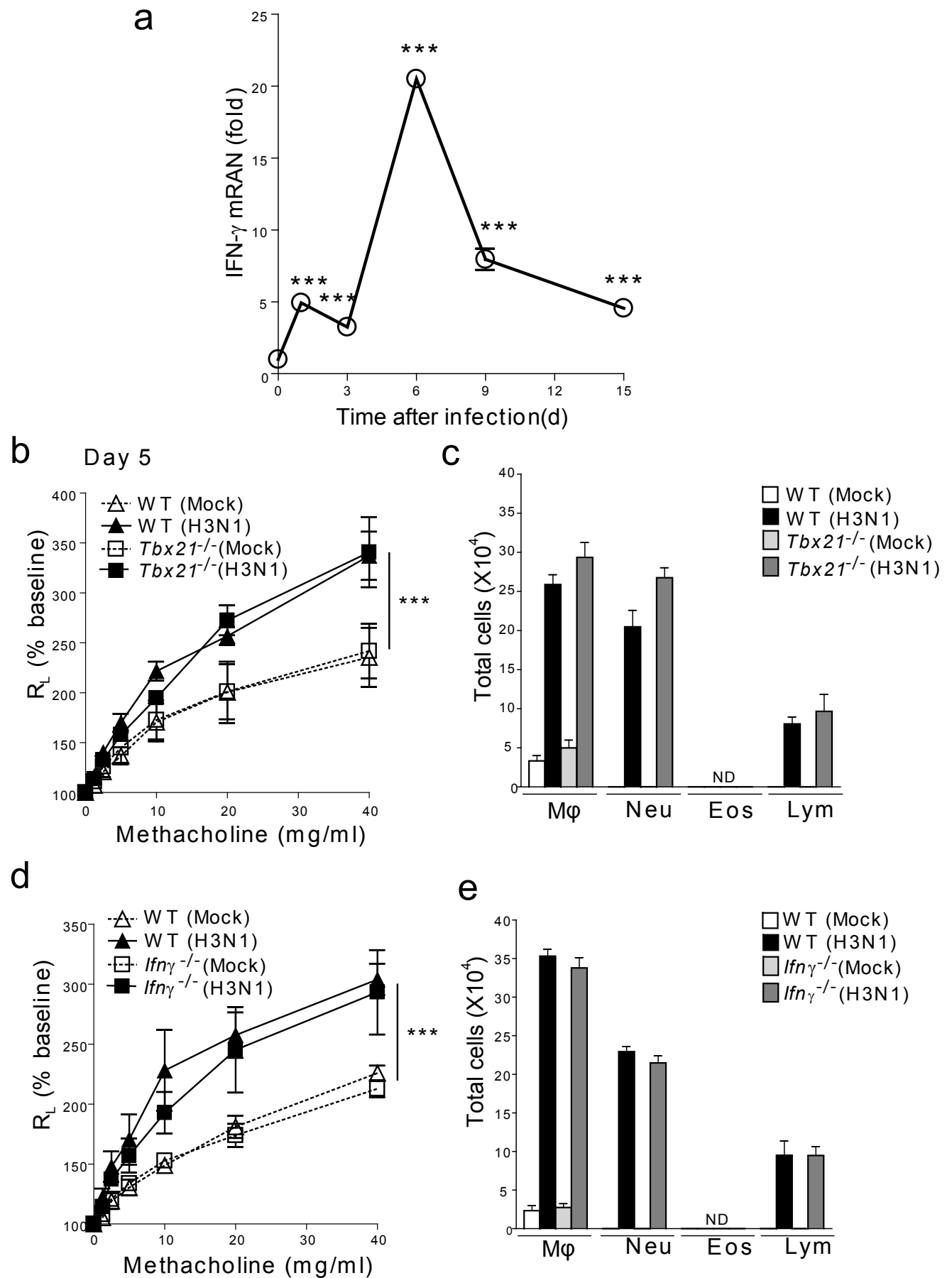




# Sup Fig 5

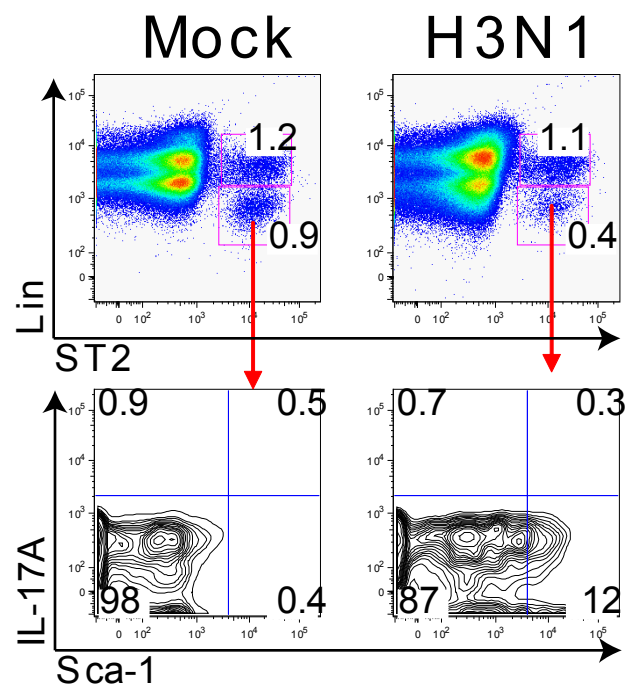


# Sup Fig 6

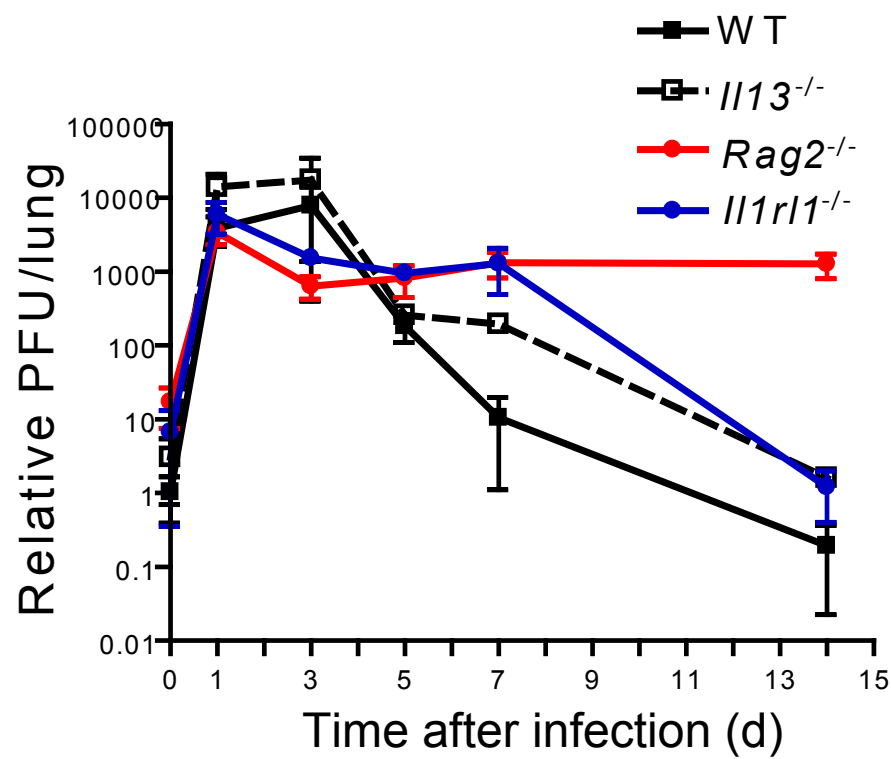


# Sup Fig 7

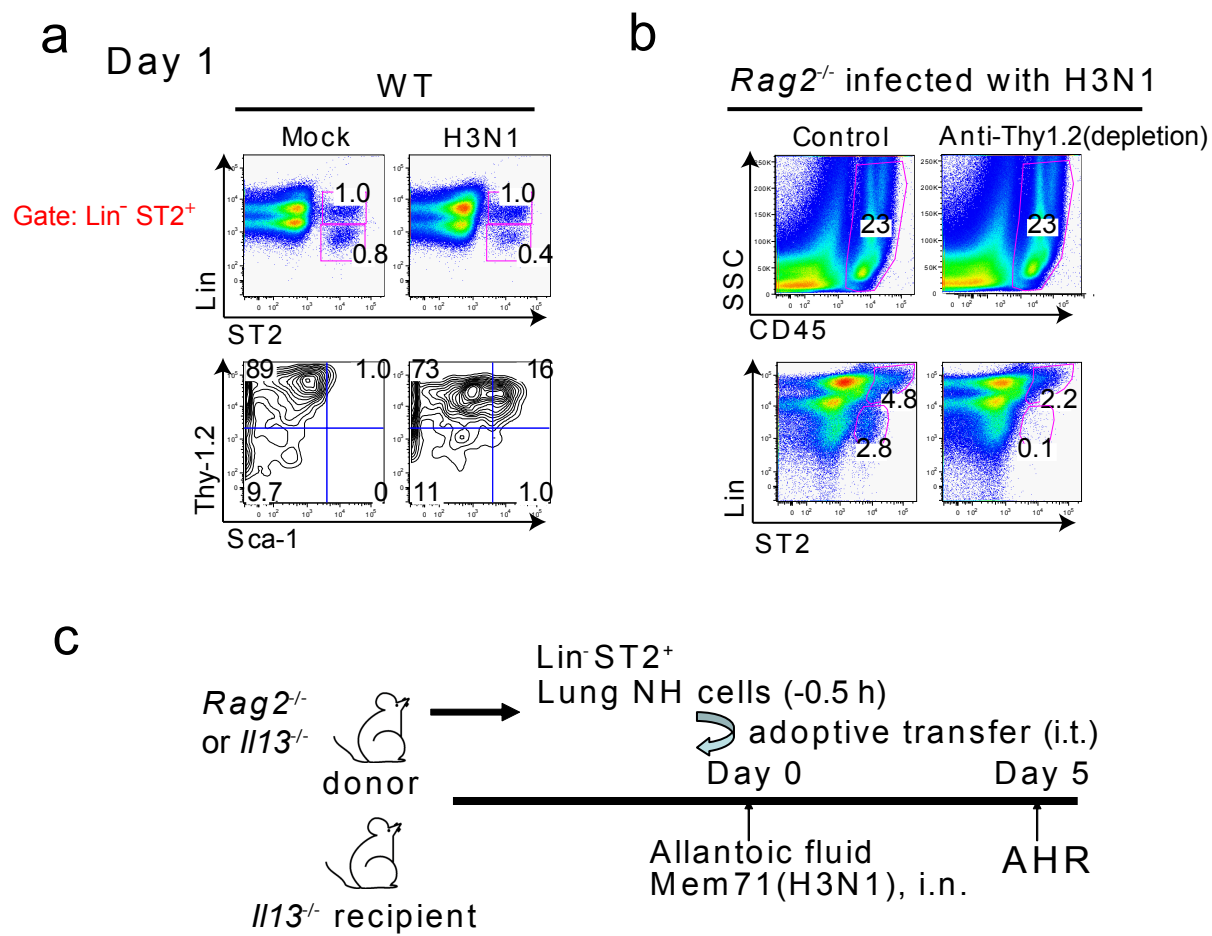
Day 1



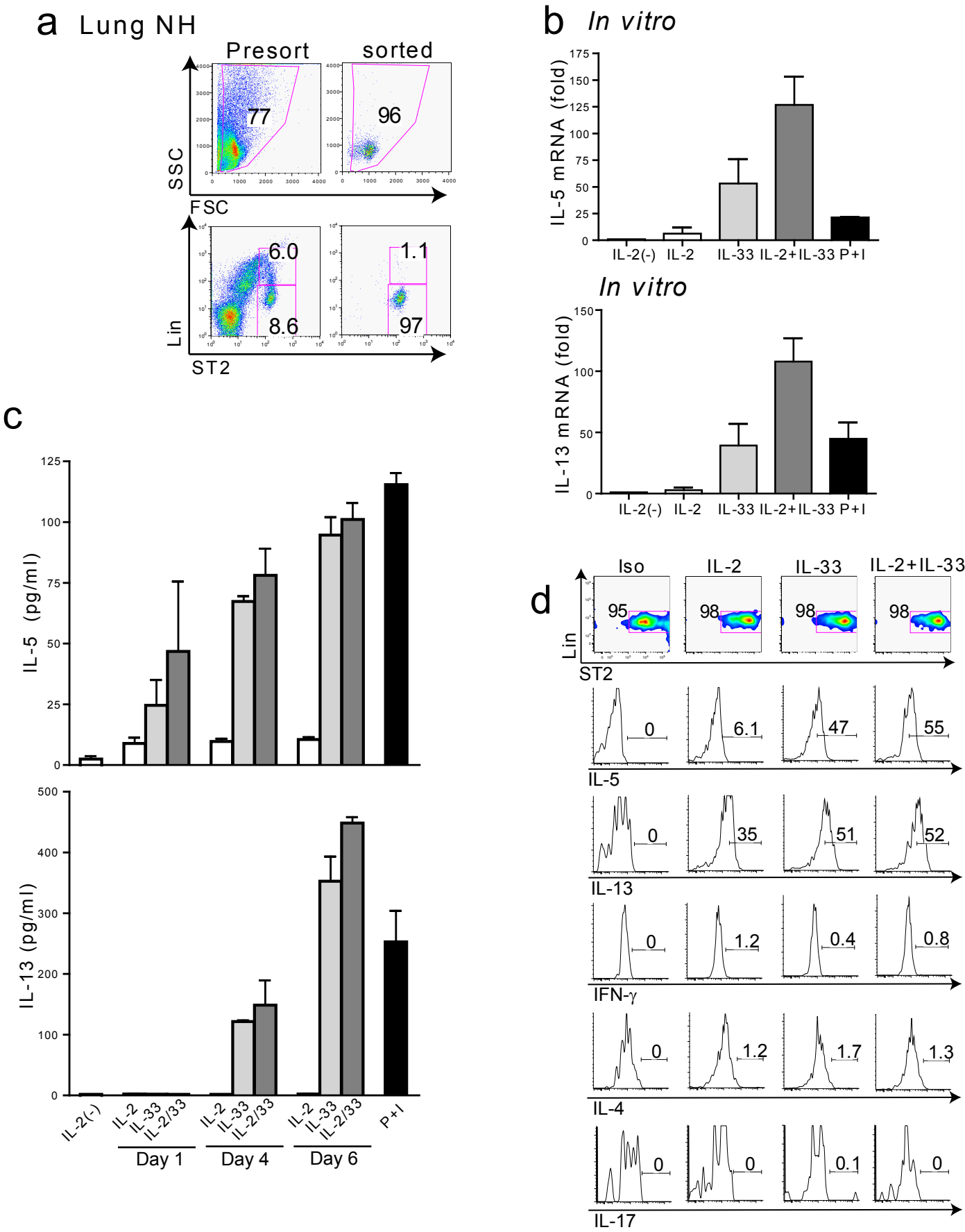
Sup Fig 8



# Sup Fig 9



# Sup Fig 10



Sup Fig 11

